SOYBEAN GROWTH RESPONSE TO WATER SUPPLY AND ATMOSPHERIC CARBON DIOXIDE ENRICHMENT

Stephen A. Prior and Hugo H. Rogers

National Soil Dynamics Laboratory, Agricultural Research Service, U.S. Department of Agriculture, P.O. Box 3439, Auburn, AL 36831-3439

ABSTRACT: Growth response of soybean [Glycine max (L.) Merr. 'Bragg'] grown in open top field chambers at five carbon dioxide (CO₂) concentrations ranging from 349 to 946 µLL-1 and under two water regimes was examined. During reproductive growth, plants grown under CO₂ enrichment exhibited increases in total leaf area and dry weight. Water stress inhibited growth at all CO₂ levels, but the relative enhancement of growth due to CO2 enrichment under water-stressed (WS) conditions was greater than under well-watered (WW) conditions. Water-stressed plants grown under 946 μLL⁻¹ CO₂ were larger than WW plants grown under 349 µLL-1 CO. Reproductive yield increases were represented by increases in seed number rather than larger seeds. Although water stress reduced yield, the relative increase in seed number in response to elevated CO₂ was greater for WS plants. Leaf tissue analysis suggested that a phosphorus deficiency may have restricted the seed dry weight response to elevated CO₂. The mean relative growth rate (RGR) and mean net assimilation rate (NAR) increased with CO₂ concentration in the first interval (5 to 14 days after planting) and diminished with time thereafter for each CO, level. At the second interval (14 to 63 days), the direct effect of NAR was offset by lower leaf area ratio (LAR). However, the LAR was greater for WS plants but the response of RGR to CO₂ was similar under both water treatments. At the third interval (63 to 98 days), the RGR for WS plants remained constant across CO₂ treatments, whereas under WW conditions a level response of NAR coupled with a negative response of LAR resulted in a decrease in RGR under CO₂-enriched conditions. The decrease in

LAR was attributed to a decrease in specific leaf area. Leaf weight ratio was unaffected by CO₂.

INTRODUCTION

Global atmospheric CO₂ concentration is increasing (Keeling et al., 1989). Since plants respond directly to CO₂, it is important to understand how this elevated concentration will directly affect the productivity of agricultural ecosystems. Carbon dioxide enrichment frequently increases net photosynthesis, plant water use efficiency, and biomass production and yield (Rogers et al., 1993). However, in order to make reliable predictions of future crop response to elevated CO₂ concentration, it will be necessary to consider the interactions of CO₂ with other environmental factors.

Chief among these is available soil water which is a major limitation to crop production (Boyer, 1982). Annual yield fluctuations and the occurrence of water stress at critical stages of growth have been largely attributed to variations in the amount and distribution of precipitation (Turner and Kramer, 1980). Previous work has suggested potential amelioration of plant water stress by increased atmospheric CO₂ level due to partial stomatal closure (Rogers et al., 1984b; Prior et al., 1991). Reports that increased CO₂ ameliorates water stress are not uncommon (Acock and Allen, 1985; Rogers and Dahlman, 1993). Controlled environment studies have demonstrated that although water stress greatly reduces plant growth, the relative increase in total dry weight and yield of water-stressed plants in response to CO₂ enrichment were as great as under well-watered conditions (Gifford, 1979; Sionit et al., 1980; Sionit et al., 1981).

The purpose of the present experiment was to investigate the long-term effects of elevated CO₂ and water supply on soybean growth, development, and yield. Soybean plants were grown in large containers in open top chambers (Rogers et al., 1983b) under various CO₂ concentrations. Plants were subjected to repeated drying cycles for most of the vegetative period (except germination and early seedling phase) and all of the reproductive stage.

MATERIALS AND METHODS

Soybean plants were grown in open top field chambers at different atmospheric CO₂ concentrations and in open plots (no chambers) under ambient atmospheric

conditions. There were two replicates of six CO_2 treatments which had a mean seasonal daytime CO_2 concentration of 348 ± 19 (ambient plots without chambers), 349 ± 19 (ambient chamber), 421 ± 22 , 496 ± 24 , 645 ± 33 and 946 ± 49 µLL⁻¹. Description of the CO_2 exposure system has been reported elsewhere (Rogers et al., 1983b; Prior et al., 1991)

Seeds were inoculated (Nitragin Co.)¹ and planted in 16.5-L pots containing 13 L of a 2:1:1 mixture by volume of sandy clay loam soil:sand:Metro-Mix 220 (W.R. Grace Co.). Plants were thinned for uniformity at 5, 10, and 14 days after planting, to a final density of one plant per pot. Each study plot contained 20 plants which were divided into two groups of 10 and subjected to two watering regimes. The experiment was a split plot design with a randomized complete block arrangement of the main-plot factor (CO₂) treatment) with two blocks. The treatments of the second factor (water treatment) were randomly assigned to subplots (pots) within each main plot. Differential water treatments were established by using vacuum gauge tensiometers and by erecting removable transparent plastic covering within all plots during periods of rainfall. Pots were placed on racks to insure proper drainage. Plants were rewatered when tensiometers reached -0.005 to -0.015 MPa and -0.075 to -0.085 MPa for well-watered controls (WW) and water-stressed (WS) plants, respectively. Drying cycles started two weeks after emergence when plants were at the third node stage (V-3; Ferr and Caviness, 1977). One liter of a N-free nutrient solution (Israel, 1981) per pot was added once a week for the duration of the growing season.

Plants were harvested at 14, 63, 98, and 140 days after planting corresponding to the seedling, anthesis, pod fill, and maturity stages. At each harvest, six randomly chosen plants were collected from each water by CO₂ treatment combination. Leaf area was determined photometrically and dry weights of plant parts were determined after drying at 55°C for three days. In addition, the following growth parameters were determined using classical growth analysis techniques (Kvet et al., 1971): (1) relative growth rate (RGR), the amount of dry matter produced per unit of dry matter present per unit time

¹Trade names and products are mentioned solely for information. No endorsement by the USDA is implied.

(kg kg⁻¹ day⁻¹); (2) net assimilation rate (NAR), the amount of dry matter produced per unit leaf area per unit time (g m⁻² day⁻¹); (3) leaf area ratio (LAR), the amount of leaf area per total plant dry weight (m² kg⁻¹); (4) specific leaf area (SLA), the amount of leaf area per total leaf dry weight (m² kg⁻¹); and (5) leaf weight ratio (LWR), the amount of leaf dry weight per total plant dry weight (kg kg⁻¹). Mean RGR and NAR were calculated for three intervals: 5 to 14 days, 14 to 63 days and 63 to 98 days. For the first interval calculations, the plant dry weight at day 5 averaged 0.144 g (mean dry weight per seed at planting) and the cotyledonary photosynthetic area averaged 3.5 cm².

Statistical analysis of data were performed using standard analysis of variance (ANOVA) and regression techniques (Statistical Analysis Systems, SAS Institute, Cary, NC).

RESULTS AND DISCUSSION

Growth, Development, and Yield

At 14 days after planting, low soil water availability had no effect on measured variables and therefore analyses were performed on combined WS and WW data (Table 1). The expected increase in total dry weight and leaf area due to CO₂ enrichment as reported by others (Rogers et al., 1984a; Patterson and Flint, 1980) was not found; lack of differences may be attributed to competition from other plants since a final density of one plant per pot was not attained until this harvest. Also growth from cotyledonary food supplies could dampen the CO₂ effect.

At anthesis (63 days) and pod fill (98 days), total dry weight under both water regimes was enhanced by the higher concentrations of CO₂ (Tables 2 and 3). At anthesis, dry weights of plants for the WW and WS treatments were ~88% higher when grown at 946 μLL⁻¹ vs 349 μLL⁻¹ CO₂, while at pod fill they were 32% and 93% higher, respectively. These results are in general agreement with data from other CO₂ studies with soybean of similar age grown under well-watered conditions (Patterson and Flint, 1980; Patterson and Flint, 1982; Rogers et al., 1983a; Rogers et al., 1984a)

At anthesis, water stress reduced total plant dry weight relative to WW control proportionately more at high CO₂ than at ambient CO₂ (Table 2). The opposite was observed at pod fill (Table 3). Despite this adverse stress effect, relative increases in total dry weight were as great or greater for water stressed plants grown under CO₂ enrichment

Table 1. Mean total dry weight, leaf area, and height for soybeans grown under two water regimes in open plots and five CO₂ treatments within chambers at 14 days after planting (well-watered = WW, water-stressed = WS).

CO ₂ (μLL ⁻¹)	Drv Wt. (g)		Leaf Ar	Leaf Area (cm²)		Height (cm)	
	ww	WS	. WW	WS	ww	WS	
348‡	0.41	0.39	43	43	6.5	6.5	
349	0.39	0.36	54	50	6.9	6.8	
421	0.41	0.42	55	53	6.7	7.2	
496	0.41	0.46	53	57	7.0	7.2	
645	0.44	0.44	54	51	6.9	7.4	
946	0.51	0.44	60	55	7.2	7.0	
s _x ¶	0.0	04	3.	3.5		2	
CV (%)	10	.8	. 10	10.6		7.8	
b _o [§]	0.4	12	52	52.3		6.9	
b _{chamber}	NS		N	NS		NS	
b _{linear}	NS		N	S	NS		
bquadratic	NS		N:	S	NS		
R²	NS		N:	NS		NS	

[‡] Ambient plot (no chamber).

which are in agreement with results reported for wheat (*Triticum aestivum* L.) by Gifford (1979). Others have reported that wheat plants subjected to acute water stress during early reproductive growth exhibit increases in total dry weight due to additional CO₂; however, total dry weights were reduced about equally by stress under both low and high CO₂ concentrations (Sionit et al., 1980; 1981).

Effects of CO₂ enrichment on plant height at anthesis and pod fill are shown in Tables 2 and 3. Height differences, while statistically significant, were not great at anthesis; the effect disappeared by pod fill. Height at both sampling times was unaffected by stress.

 $[\]P$ $\mbox{ s}_x$ and CV (%) from ANOVA.

[§] Curves were fitted against actual CO₂ values. Note: Linear and quadratic coefficients and their standard errors should be multiplied by 10⁻³.

Table 2. Mean total dry weight, leaf area, and height for soybeans grown	
two water regimes in open plots and five CO ₂ treatments within chambers :	at 63
days after planting (well-watered = WW, water-stressed = WS).	

CO ₂ (μLL ⁻¹)	Drv Wt. (g)		Leaf Ar	Leaf Area (cm²)		Height (cm)	
	WW	WS	WW	WS	WW	WS	
348‡	34.3	28.9	3820	3034	60	50	
349	46.5	27.0	5150	3429	77	73	
421	53.5	35.1	5425	4299	78	80	
496	61.8	38.6	5743	4092	. 78	78	
645	72.4	50.2	6060	5268	84	87	
946	87.6	51.0	6582	5140	81	80	
s _x ¶	2.	4	1.2		2.4		
CV (%)	11	.7	15	.9	8.0		
b _o §	10.8	-23.8	3188.7	1934.6	35.	.1	
$b_{chamber}$	15.0 [†]	-1.8	1136.6 [†]		14.	5 [†]	
blinear	67.5	190.7	2486.9		93.6		
bquadratic	NS	-0.1	NS		-0.06		
R ²	0.99	0.99	0.97		0.9	8	

[‡] Ambient plot (no chamber).

Leaf area at anthesis increased with CO₂ enrichment in both water treatments (Table 2). There was no stress by CO₂ treatment interaction; plants in both water treatments responded to elevated CO₂ similarly. At pod fill we observed a stress by CO₂ treatment interaction (Table 3). Despite a reduction in leaf area due to water stress, the proportional increase in leaf area due to CO₂ enrichment was greater for WS plants since additional CO₂ had no significant effect on leaf area of WW plants. This interaction could have been the result of less rapid rate of leaf expansion in WS plants compared to WW plants between 63 and 98 days after planting under CO₂ enrichment. The WW

 $[\]P$ $\mbox{ s}_{x}$ and CV (%) from ANOVA.

[§] Curves were fitted against actual CO₂ values. Note: Linear and quadratic coefficients should be multiplied by 10⁻³.

[†] Significant F (0.95 level) for chamber effect.

Table 3. Mean total dry weight, leaf area, and pod dry weight for soybeans grown under two water regimes in open plots and five CO₂ treatments within chambers at 98 days after planting (well-watered = WW, water-stressed = WS).

CO ₂ (μLL ⁻¹)	Drv Wt. (₫)		Leaf Area (cm²)		Pod Drv Wt. (g)	
	WW	WS	WW	WS	WW	WS
348‡	131.4	101.5	7873	6327	39.3	31.4
349	181.2	102.8	9365	7182	47.5	30.3
421	230.0	122.0	11114	8231	53.5	33.4
496	204.4	142.6	9931	8860	51.1	35.8
645	228.9	168.9	10476	9611	52.1	42.6
946	239.5	198.8	9939	9468	68.6	55.8
$S_N^{\frac{4}{4}}$	9.3		445.1		2.6	
CV (%)	7.	9	7.8		7.7	
b₀ [§]	108.2	45.0	7872.8	98.8	29.1	15.1
$b_{chamber}$	39.	8 [†]	2292.2 [†]	940.3 [†]	3.3	
b_{linear}	114	.6	NS	23139.2	38	.1
bquadratic	N:	S	NS	-15.1	N:	S
R ²	0.9	7	NS	0.99	0.9	9

[‡] Ambient plot (no chamber).

plants may have reached their maximum leaf area before 14 weeks whereas leaf area expansion for WS plants continued until week 14.

The higher CO₂ concentration delayed anthesis (and shortened its duration) by about 2 days under both water treatments. Other investigators have reported a delay in anthesis for soybean due to CO₂ enrichment (Hesketh and Hellmers, 1973; Rogers et al., 1984a). Water stress had little effect on the timing of anthesis regardless of CO₂ level. Despite the shorter flowering period, high CO₂ grown plants probably had fewer flower abortions since increases in pod number were observed for both water treatments at pod fill (data not shown) and at maturity harvest (Table 4). The extra CO₂ extended pod

 $[\]P$ s_x and CV (%) from ANOVA.

[§] Curves were fitted against actual CO₂ values. Note: Linear and quadratic coefficients should be multiplied by 10⁻³.

[†] Significant F (0.95 level) for chamber effect.

Table 4. Mean total pod number, pod dry weight, and weight per pod for
soybeans grown under two water regimes in open plots and five CO, treatments
within chambers at maturity (well-watered = WW, water-stressed = WS).

CO ₂ (μLL ⁻¹)	Pod Number		Pod Dr	Pod Drv Wt. (g)		Wt. Per Pod	
	WW	WS	WW	WS	WW	WS	
348‡	163	128	64.4	58.2	0.40	0.46	
349	186	118	82.8	62.1	0.45	0.53	
421	226	127	101.9	73.8	0.45	0.58	
496	211	142	92.2	73.8	0.44	0.52	
645	205	156	90.3	74.8	0.44	0.48	
946	219	157	93.5	72.8	0.43	0.46	
S_N	5	.5	4.	.2	0.01		
CV (%)	7	. 1	6.	8	5	.3	
bo	153.5	34.3	64.4	58.2	0.40	0.51	
b _{chamber} *	14.8	-12.2	27.7 [†]	13.2	0.04*	0.09†	
b_{linear}	26.7	344.4	NS	NS	NS	-0.16	
bquadratic	NS	-0.2	NS	NS	NS	NS	
R ²	0.84	0.99	NS	NS	N'S	0.74	

[‡] Ambient plot (no chamber).

filling by ~3 days; the increases in pod dry weight seen under both water treatments (i.e., at the pod fill sampling) may have been due to this longer pod filling period. Duration of seed filling remained unchanged at high CO₂, but was shortened by 2 days due to water stress.

The overall effects of CO₂ and water treatment on yield components are shown in Table 3 (pod fill). Water stress reduced both pod dry weight (Table 3) and pod number (data not shown) at all CO₂ level. Increases in pod dry weight due to CO₂ enrichment were observed under both water regimes; pod dry weight increases for WW

 $[\]P$ s_x and CV (%) from ANOVA.

[§] Curves were fitted against actual CO₂ values. Note: Linear and quadratic coefficients should be multiplied by 10⁻³.

[†] Significant F (0.95 level) for chamber effect.

plants may be due to an increase in weight per pod, while more pods per plant was responsible for pod dry weight increases under WS conditions.

The number of pods produced at maturity (Table 4) was slightly greater under CO₂ enrichment. Water stress reduced pod production at maturity. However, the relative increases in pod and seed number in response to additional CO₂ were greater for plants grown under water stress. An increase in pod number resulted in a greater number of seeds per plant (Table 5), but there was only a small nonsignificant increase in pod dry weight with increasing CO₂ concentration (at both water levels). Weight per pod declined with increasing CO₂ under WS conditions but remained constant under WW conditions across the various CO₂ levels. Weight per seed increased slightly in response to water stress and decreased in response to CO₂, especially under stress conditions.

Our present observations indicate only 9% and 12% nonsignificant increases in seed dry weight for WW and WS plants, respectively, at 946 vs 349 µLL⁻¹ CO₂. Other studies have reported that CO₂ enrichment increased soybean pod number but this did not necessarily guarantee an increase in pod dry weight (Rogers et al., 1984a; 1986); CO₂ yield responses in terms of seed number and seed dry weight were not reported in these studies. Other studies have found that increases in seed yield observed under CO₂ enrichment were associated with greater numbers of seeds per plant, with seed size remaining constant or increasing slightly (Gifford, 1979; Sionit et al., 1981; Jones et al., 1984a; Rogers et al., 1983a). These different responses to CO₂ enrichment may have occurred because of differences in the availability of mineral nutrients.

In our study, the small yield response to elevated CO₂ may have been related to an inadequate supply of phosphorus. Generally, a P level of greater than 0.25% for leaves sampled at anthesis (Hanway and Olson, 1980) or early pod set (Jones et al., 1991) is sufficient. In this study, leaf analysis at anthesis showed a decrease in P concentration of ~33% at the high CO₂ level; the P concentration for WW and WS treatments were 0.24% and 0.30% at 349 µLL⁻¹ CO₂ and 0.16% and 0.20% at 946 µLL⁻¹ CO₂, respectively. Prior investigations have found that plants grown under CO₂ enrichment exhibit reductions in the concentration of nutrients in plant tissue (Conroy, 1992; Conroy et al., 1992; Porter and Grodzinski, 1989; Rogers et al., 1984a). In the present study, the application rate of only one liter per week of nutrient solution (containing 0.25 mM of P) coupled with a low P level in the soil may not have been sufficient to meet the P requirement of the

Table 5. Mean seed number, seed dry weight, and weight per seed for soybeans grown under two water regimes in open plots and five CO₂ treatments within chambers at maturity (well-watered = WW, water-stressed = WS).

CO ₂ (μLL ⁻¹)	Seed Number		Seed Dr	Seed Drv Wt. (g)		Wt. Per Seed	
	WW	WS	WW	WS	WW	WS	
348‡	319	272	46.1	40.8	0.14	0.15	
349	409	254	53.5 .	41.1	0.13	0.16	
421	478	275	65.5	50.0	0.14	0.18	
496	465	302	58.1	48.0	0.13	0.16	
645	441	341	57.4	48.2	0.14	0.14	
946	529	379	57.8	45.7	0.11	0.12	
s_x^{\P}	26	5.2	2.8		0.008		
CV (%)	11	.5	6.	6.7		8.9	
bo§	269.4	199.6	46.1	40.8	0.16	0.18	
b _{chamber}	113.5 [†]	-8.3	14.8*	5.8 [†]	-0.009 [†]	0.02†	
blincar	141.6	207.6	NS	NS	-0.04	-0.09	
bquadratic	NS	NS	NS	NS	NS	NS	
R ²	0.88	0.97	NS	NS	0.91	0.90	

[†] Ambient plot (no chamber).

larger CO₂-enriched plants. This P level may have been sufficient initially since the total pod dry weight at the pod fill sampling increased significantly with increasing CO₂ concentration (Table 3). However, as the season progressed the demand for P probably increased and altered assimilate translocation to the seeds, thus resulting in the small nonsignificant seed yield increase observed at maturity (Table 5). Studies conducted with soybeans grown under ambient CO₂ concentration and various levels of phosphorus demonstrated that the P level used in our study (0.25 mM) was in the low to intermediate range in terms of yield response (Israel, 1987). Giaquinta et al. (1981) reported that

[¶] s_x and CV (%) from ANOVA.

[§] Curves were fitted against actual CO₂ values. Note: Linear and quadratic coefficients should be multiplied by 10⁻³.

[†] Significant F (0.95 level) for chamber effect.

increasing the nutrient phosphate concentration decreased the starch/sucrose ratio which in turn increased the dry matter accumulation in the seeds. Cure et al. (1988) suggested that under P limiting conditions, N₂-fixing plants may not be as responsive to extra CO₂ with respect to growth and yield as plants dependent on inorganic sources of N.

In contrast to results reported here, greater yield increases due to CO₂ were reported for those studies which had more frequent applications of nutrient solution (Sionit et al., 1981), adequate rates of fertilizer applied to the soil (Gifford, 1979; Jones et al., 1984; Rogers et al., 1984a), and unrestricted soil volume for root development and exploration of available nutrients (Jones et al., 1984; Rogers et al., 1986). The discrepancies between these findings and ours indicate the importance of considering fertility requirements of crops in a CO₂-enriched environment.

Another important factor which may have contributed to the small CO₂ yield response observed in our study could be related to root restriction which may have limited photosynthetic capacity of pot grown plants (i.e., sink limited feedback inhibition) (Sionit et al., 1984; Arp, 1991; Thomas and Strain, 1991). Root restriction within field soil profiles can also occur (e.g., traffic induced soil compaction, tillage and natural hardpans, etc.). Therefore, results of containerized studies may in some cases reflect responses of plants growing in the field. This suggests a need to conduct CO₂ studies in the field under different tillage systems and with different soil series.

The harvest index decreased with elevated CO₂ and increased under chronic water stress conditions (Table 6). Seed oil content was observed to increase slightly with increasing CO₂. Elevated CO₂ had no effect on the fatty acid composition of the seed oil (data not shown). There was a trend toward decreasing seed protein value with increasing CO₂. These findings are in general agreement with observations reported by Rogers et al. (1984a). In the present study, water stress had no significant effect on either seed oil or protein values.

Despite the small yield response to CO_2 , increased total dry matter production due to CO_2 under both water regimes represents more carbon fixed and in turn more carbon input to the soil. This input may influence carbon storage/cycling as well as belowground physical and biological processes. More dry matter returned to the soil surface may also be important for erosion control in light of the increasing adoption of conservation tillage. At present, CO_2 field studies have only begun to consider these topics.

Table 6. Mean total dry weight, harvest index, and total oil (%) for soybeans
grown under two water regimes in open plots and five CO2 treatments within
chambers at maturity (well-watered = WW, water-stressed = WS).

CO ₂ (μ L L ⁻¹)	Dry Wt. (g)		<u>Harve</u>	<u>Harvest Index</u>		Total Oil (%)	
	WW	WS	WW	WS	WW	WS	
348‡	91.1	79.8	0.57	0.56	20.4	20.5	
349	128.4	92.6	0.49	0.53.	21.2	20.7	
421	157.5	104.0	0.48	0.54	21.8	21.4	
496	148.9	106.1	0.46	0.49	22.0	21.5	
645	156.8	117.6	0.43	0.49	20.9	21.4	
946	156.3	117.0	0.43	0.45	21.1	21.0	
s_x^{\P}	7	7.4 0		006	0.16		
CV (%)	5	.8	2	.7	2.1		
b ₀ [§] .	80.7	67.3	0.70	0.67	19	.2	
b _{chamber}	51.9 [†]	20.3*	~0.08 [†]	-0.03*	0.80†		
b _{linear}	29.7	36.0	-0.48	-0.36	5.1		
bquadratic	NS	NS	0.003	0.0002	-0.0043		
R ²	0.94	0.93	0.99	0.97	0.83		

i Ambient plot (no chamber).

Growth Analysis

The net assimilation rate (NAR) and relative growth rate (RGR) over three time intervals (5 to 14 days, 14 to 63 days and 63 to 98 days) during the growing season are given in Fig. 1. Data shown for the initial interval are mean values under WW conditions since no water stress had been imposed during this period, whereas the latter two intervals include data for both WW and WS treatments.

The effects of CO₂ enrichment on NAR were dependent on plant age, length of exposure, and water treatment (Fig. 1A). During the first interval (5 to 14 days), NAR

 $[\]P$ s_x and CV (%) from ANOVA.

[§] Curves were fitted against actual CO, values. Note: Linear and quadratic coefficients should be multiplied by 10⁻³.

[†] Significant F (0.95 level) for chamber effect.

(an approximate measure of leaf C-assimilatory capacity) was observed to increase with increasing atmospheric CO₂ concentration. The highest value of NAR (19.14 g m² day⁻¹ at 946 μLL⁻¹ CO₂) was 33% greater compared to the 349 μLL⁻¹ CO₂ treatment level. In another open top chamber study, soybean sampled over a similar time interval showed an NAR increase of 67% due to CO₂ enrichment (Rogers et al., 1984a). Growth chamber CO₂ studies conducted with wheat showed similar changes in NAR (Neales and Nicholls, 1978; Sionit et al., 1979).

In this study, the maximum NAR enhancement due to CO_2 enrichment was observed during the second interval (weeks 2-9). Net assimilation rate increased 53% at the 946 μ LL⁻¹ CO_2 under WW conditions. Although water stress reduced NAR more at higher CO_2 levels (645 to 946 μ LL⁻¹ CO_2), NAR was increased by ~35% at these CO_2 treatments relative to ambient conditions. By the third interval (weeks 9-14) NAR was relatively constant across CO_2 treatments under WW conditions. The highest value of NAR for WS plants (3.4 g m² day -1 at 946 μ LL-1 CO_2) was 46% greater than at 349 μ LL-1 CO_2 .

Increasing the CO₂ concentration had the greatest effect on RGR during the first interval (Fig. 1B). Initial increases in RGR under elevated CO₂ were attributed to increased NAR (Fig. 1A). As with NAR, RGR decreased at each CO₂ value as the plants aged, especially between the second and third interval. The initially high values of RGR and NAR observed during earlier phases of growth followed by a gradual decline with plant age (ontogenetic drift; Kvet et al., 1971) are consistent with those reported for soybean under ambient CO₂ conditions (Koller et al., 1970).

Despite increased NAR due to higher CO₂ during the second interval, RGR values for both WW and WS plants were only slightly higher under higher CO₂. RGR under WS conditions was equally reduced at the various CO₂ concentrations, thus the CO₂ response was similar to that observed under WW conditions.

For the third interval, we detected a decrease in RGR under WW conditions, whereas WS values for RGR remained fairly constant despite increasing CO₂ concentration. The greatest reduction of RGR for WW plants (41%) was observed when the CO₂ concentration was increased from 349 to 946 µLL⁻¹. Several investigators have reported that both RGR and NAR were initially increased during early development but declined as plants grew older, especially with further exposure to elevated CO₂ (Huges

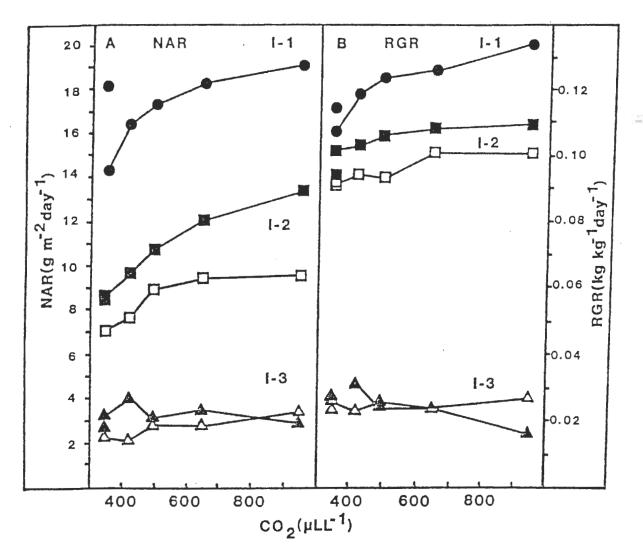


FIGURE 1. Mean NAR and RGR for well-watered (closed symbols) and water-stressed (open symbols) soybean grown in open plots (lone symbols) and five CO_2 treatments within chambers (connected symbols) at three time intervals [II = 5 to 14 days (o, •); I2 = 14 to 63 days (\square , \square); I3 = 63 to 98 days (\triangle , \triangle) after planting].

and Freeman, 1967; Hurd, 1968; Huges and Cockshull, 1969; Neales and Nicholls, 1978; Rogers et al., 1984a; Tolley and Strain, 1984).

The effects of CO₂ enrichment and water treatment on leaf area ratio (LAR) and its two components, specific leaf area (SLA) and leaf weight ratio (LWR), are shown in Figure 2. Since the stress treatment was not initiated until 2 weeks (after emergence), data for this period represent WW mean values only (Fig. 2A). In general, the distribution of plant dry matter to photosynthetically active tissue (LAR) declined with time and values of LAR at 349 µLL⁻¹ CO₂ were invariably greater than those found at

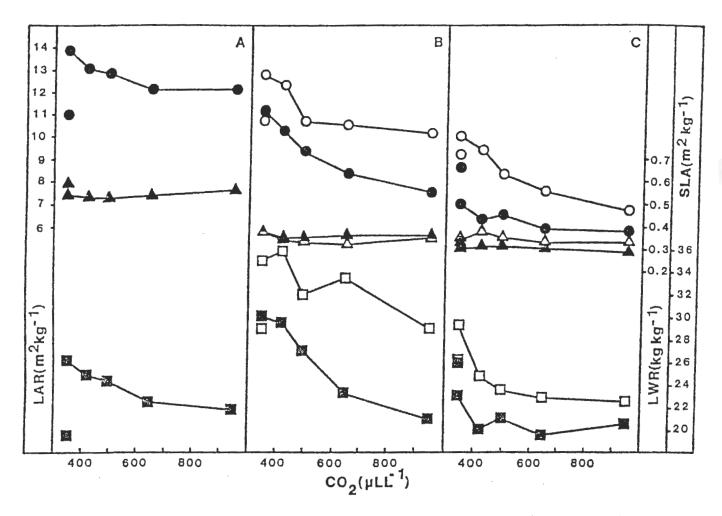


FIGURE 2. LAR $(0, \bullet)$, SLA (\square, \blacksquare) and LWR $(\triangle, \blacktriangle)$ for well-watered (closed symbols) and water-stressed (open symbols) soybean grown in open plots (lone symbols) and five CO_2 treatments within chambers (connected symbols) at 14 (A), 63 (B) and 98 (C) days after planting.

increased levels of CO₂. As reported by other investigators for several species (Huges and Freeman, 1967; Hurd, 1968; Neales and Nichols, 1978; Patterson and Flint, 1980; Rogers et al., 1984a; Tolley and Strain, 1984), this reduction in LAR was not attributed to a change in dry matter allocation to the leaves (LWR) but to a reduction in leaf area per unit leaf dry weight (SLA). Differences in LWR were small at the various CO₂ concentrations at each harvest; LWR decreased from day 14 to 63 and then remained relatively constant thereafter. Reduction of SLA with increasing CO₂ was clearly evident at each harvest, with the highest SLA values being observed at day 63; the decrease in SLA under high CO₂ may be attributed to a greater starch accumulation in the leaves (Patterson and Flint, 1980; Madsen, 1968) and/or to an increase in leaf thickness (Hurd, 1968; Thomas and Harvey, 1983).

Although LAR was depressed by CO₂ enrichment under both water treatments, the LAR of WS plants was always higher than LAR of WW plants at each CO2 level (Fig. 2B and 2C). At day 63, over half of the decrease in LAR of WS plants occurred over the range 349 to 496 μLL⁻¹ CO₂, with the greatest decrease being observed at 946 μLL⁻¹ CO_2 (Fig. 2B). Under WW conditions, the decrease in LAR from 349 to 496 μLL^{-1} CO_2 were very similar to the WS response, and the LAR values of WS plants were about 15% greater than their WW counterparts. The greater reduction in LAR for WW plants at 645 and 946 µLL-1 CO₂ resulted in larger differences between values for WW and WS plants at these CO2 levels. At day 98, LAR of WS plants was decreased by 33% when CO2 was increased from 349 to 946 µLL-1 (Fig. 2C). Under WW conditions, reductions in LAR were less dramatic over the CO₂ concentration range. At this harvest, the greatest differences between water treatments were observed under low CO₂ and diminished when CO₂ was raised. LAR for WS plants at 349 to 421µLL⁻¹ CO₂ were about 46% greater than values for WW plants, whereas LAR at 946 µLL-1 CO₂ was only 16% greater. Reduction of SLA with increasing CO₂ was responsible for the reductions in LAR observed at each harvest. Conversely, LWR remained relatively constant with increasing CO₂. Differences due to water treatment were not great at 63 days after planting, and although LAR values for WS plants at 98 days after planting were invariably greater than values for WW plants, the differences were not large.

The observed reductions in LAR coupled with changes in NAR are important because these are the two component growth parameters of RGR (Figures 1 and 2). At the first interval, the opposing effect of LAR on NAR was not great since RGR was enhanced when the CO₂ concentration was raised from 349 to 946 μLL⁻¹. However, for the second interval, the reduction in LAR was large enough to depress RGR under both water treatments despite significant increases in NAR at high CO₂. Note that LAR offset NAR more under WW conditions, thus accounting for the similar responses of RGR to CO₂ for WW and WS plants. At the third interval, a fairly constant NAR coupled with reduced LAR resulted in the RGR for WW plants being depressed at the higher CO₂ concentrations (496 to 946 μLL⁻¹). In contrast, the RGR of WS plants remained essentially unchanged under higher CO₂ since increases in NAR were neutralized by decreases in LAR. Several investigators have reported similar changes in time for RGR through adjustments of NAR and LAR under elevated CO₂ concentration (Huges and

Freeman, 1967; Hurd, 1968; Neales and Nichols, 1978; Huges and Cockshull, 1969; Rogers et al., 1984a).

CONCLUSION

Increasing the CO₂ concentration had no effect on dry weight growth at the seedling stage. This result, contrary to the expected response, may have been due to competition since each container had more than one plant up until the 14 day after planting harvest. Thus, although NAR and RGR for the first interval increased with increasing CO₂ concentration, these increases were not as great as those frequently observed during early developmental growth. Later in the season (anthesis and pod fill samplings), CO₂ enrichment did significantly increase total dry weight. Water stress inhibited growth at all CO₂ levels at both periods. However, the relative enhancement of growth due to increasing CO₂ concentration under WS conditions was as great or greater than that under WW conditions. At each of these periods, increasing the CO₂ concentration was observed to have a compensating effect for water stress. For both periods, WS plants grown under 946 µLL⁻¹ CO₂ were larger than WW plants grown under 349 µLL⁻¹ CO₂. This compensating effect was due to the fact that WS plants under high CO₂ had growth characteristics (i.e., RGR, NAR, and LAR) similar to WW plants under ambient CO₂.

Reproductive yield increases due to CO₂ enrichment were represented by a significant rise in seed number. Although water stress had an adverse effect on yield, the relative increase in seed number in response to elevated CO₂ was greater for WS plants. There was a significant decrease in weight per seed with CO₂ under both water treatments. There was only a trend toward increase in total seed dry weight. This leads us to suggest that a limiting P supply and/or a restricting soil environment may have led to the lack of a significant seed yield response to increasing CO₂. Further research with crops grown in the field using different irrigation, fertility and tillage management practices, and different soils is needed if we are to develop a reliable knowledge base that will allow prediction of crop response in a future higher CO₂ world.

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REFERENCES:

- Arp, W. J. 1991. Effects of source-sink relations on photosynthetic acclimation to elevated CO₂. Plant, Cell and Environ. 14:869-975.
- Boyer, J. S. 1982. Plant productivity and environment. Sci. 218:443-448.
- Conroy, J. P. 1992. Influence of elevated atmospheric CO₂ concentration on plant nutrition. Aust. J. Bot. 40:445-456.
- Conroy, J. P., P. J. Milham, and E. W. Barlow. 1992. Effect of nitrogen and phosphorus availability on the growth response of *Eucalyptus grandis* to high CO₂. Plant Cell and Environ. 15:843-847.
- Cure, J. D., T. W. Rufty, Jr., and D. W. Israel. 1988. Phosphorus stress effects on growth and seed yield responses of nonnodulated soybean to elevated carbon dioxide. Agron. J. 80:897-902.
- Fehr, W. R., and C. E. Caviness. 1977. Stages of soybean development. Iowa Agric. Home Econ. Exp. Stn., Iowa Coop. Ext. Serv. Spec. Rep. 80.
- Giaquinta, R. T., B. Quebedeaux, and V. Wittenbach. 1981. Alterations in photosynthesis and assimilate partitioning between starch and sucrose in soybean leaves during seed filling, pp.549-550. IN: G. Akoyunoglou (ed.) Photosynthesis IV. Regulation of carbon metabolism. Balaban International Science Services, Philadelphia, PA.
- Gifford, R. M. 1979. Growth and yield of CO₂-enriched wheat under water limited conditions. Aust. J. Plant Physiol. 6:367-378.
- Hanway, J. J., and R. A. Olson. 1980. Phosphate nutrition of corn, sorghum, soybeans and small grains, pp. 681-691. IN: F. E. Kahasawneh, E. C. Sample, and E.J. Kamprath (eds.) Role of Phosphorus in Agriculture. American Society of Agronomy, Madison, WI.
- Hesketh, J.D., and H. Hellmers. 1973. Floral initiation in four plant species growing in CO₂-enriched air. Environ. Control Biol. 11:51-53.
- Huges, A.P., and K.E. Cockshull. 1968. Effects of carbon dioxide concentration on the growth of *Callistephus chinensis* cultivar Johannistag. Ann. Bot. 33:351-365.
- Huges, A.P., and P.R. Freeman. 1967. Growth analysis using frequent small harvests.

 J. Appl. Ecol. 4:553-560.
- Hurd, R.G. 1968. Effects of CO₂ enrichment on the growth of the young tomato plants in low light. Ann. Bot. 32:531-542.

- Israel, D. W. 1981. Cultivar and rhizobium strain effects on nitrogen fixation and remobilization by soybeans. Agron. J. 73:509-516.
- Israel, D. W. 1987. Investigation of the role of phosphorus in symbiotic dinitrogen fixation. Plant. Physiol. 84:835-840.
- Jones, P., L. H. Allen, Jr., J. W. Jones, K. T. Boote, W. J. Campbell. 1984. Soybean canopy growth, photosynthesis, and transpiration responses to whole-season carbon dioxide enrichment. Agron. J. 76:633-637.
- Jones, J. B., B. Wolf, and H. A. Mills. 1991. Plant Analysis Handbook. Micro-Macro Publishing, Inc., Athens, GA.
- Keeling, C. D., R. B. Bacastow, A. F. Carter, S. C. Piper, T. P. Whorf, M. Heimann, W. G. Mook, and H. Roeloffzen. 1989. A three dimensional model of atmospheric CO₂ transport based on observed winds: Observational data and preliminary analysis. <u>IN</u>: Aspects of climate variability in the pacific and the western americas. Volume 55. American Geophysical Union.
- Koller, H. R., W. E. Nyquist, and I. S. Chorush. 1970. Growth analysis of the soybean community. Crop Sci. 10:407-412.
- Kvet, J., P. G. Jarvis, J. P. Ondok, and J. Necas. 1971. Methods of growth analysis. IN: Catsky and P. G. Jarvis (eds.) Plant Photosynthetic Production Manual of Methods. Dr. W. Junk, The Hague, The Netherlands.
- Lemon, E. R. (ed.). 1983. CO₂ and plants: The response of plants to rising levels of atmospheric carbon dioxide. AAAS Selected Symposium 84. Westview Press, Boulder, CO.
- Madsen, E. 1968. Effect of CO₂ concentration on the accumulation of starch and sugar in tomato leaves. Physiol. Plant. 21:168-175.
- Neales, T. F., and A. O. Nicholls. 1978. Growth responses of young wheat plants to a range of ambient CO₂ levels. Aust. J. Plant Physiol. 5:45-59.
- Patterson, D. T., and E. P. Flint. 1980. Potential effects of global atmospheric CO₂ enrichment to the growth and competitiveness of C₃ and C₄ weed and crop plants. Weeds Sci. 28:71-75.
- Patterson, D. T., and E. P. Flint. 1982. Interacting effects of CO₂ and nutrient concentration. Weed Sci. 30:389-394.
- Porter, M. A., and B. Grodzinski. 1989. Growth of bean in high CO₂: Effects on shoot mineral composition. J. Plant Nut. 12(2):129-144.
- Prior, S. A., H. H. Rogers, N. Sionit, and R. P. Patterson. 1991. Effects of elevated atmospheric CO₂ on water relations of soya bean. Agric. Ecosys. Environ. 35:13-25.

- Rogers, H. H., G. E. Bingham, J. D. Cure, J. M. Smith, and K. A. Surano. 1983a. Responses of selected plant species to elevated CO₂ in the field. J. Environ. Qual. 12:569-574.
- Rogers, H. H., W. W. Heck, and A. S. Heagle. 1983b. A field technique for the study of plant responses to elevated carbon dioxide concentrations. J. Air Pollut. Control Assoc. 33:42-44.
- Rogers, H. H., J. D. Cure, J. F. Thomas, and J. M. Smith. 1984a. Influence of elevated CO₂ on growth of soybeans. Crop Sci. 24:361-366.
- Rogers, H. H., N. Sionit, J. D. Cure, J. M. Smith, and G. E. Bingham. 1984b. Influence of elevated carbon dioxide on water relations of soybeans. Plant Physiol. 74:233-238.
- Rogers, H. H., J. D. Cure, and J. M. Smith. 1986. Soybean growth and yield response to elevated carbon dioxide. Agric. Ecosys. Environ. 16:113-128.
- Rogers, H. H., and R. C. Dahlman. 1993. Crop responses to CO₂ enrichment. Vegetatio 104/105:117-131.
- Sionit, N, H. H. Rogers, G. E. Bingham, and B. R. Strain. 1984. Photosynthesis and stomatal conductance with CO₂-enrichment of container- and field-grown soybeans. Agron. J. 76:447-451.
- Sionit, N., H. Hellmers, and B. R. Strain. 1980. Growth and yield of wheat under CO₂ enrichment and water stress. Crop Sci. 20:687-690.
- Sionit, N., B. R. Strain, and H. Hellmers. 1979. Effects of different concentrations of atmospheric CO₂ on growth and yield components of wheat. J. Agric. Sci. 79:335-339.
- Sionit, N., B. R. Strain, H. Hellmers, and P. J. Kramer. 1981. Effects of atmospheric CO₂ concentration and water stress on water relations of wheat. Bot. Gaz. 142:191-196.
- Thomas, J. F., and C. N. Harvey. 1983. Leaf anatomy of four species grown under continuous long-term CO₂ enrichment. Bot. Gaz. 144:303-309.
- Thomas, R. R., and Strain, B. R. 1991 Root restriction as a factor in photosynthetic acclimation of cotton seedlings grown in elevated carbon dioxide. Plant Physiol. 96:627-634.
- Tolley, L. C., and B. R. Strain. 1984. Effects of CO₂ enrichment on growth of *Liquidambar styraciflua* and *Pinus taeda* seedlings under different irradiance levels. Can. J. For. Res. 14:343-350.
- Turner, N. C., and P. J. Kramer. (eds.). 1980. Adaptation of Plants to Water and High Temperature Stress. Wiley, New York, NY.